

TMP (Trimethoprim) ELISA Kit

Catalog No: E-FS-E020

96T/96T*3

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017

Email: techsupport@elabscience.com

Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

Test principle

This kit uses Competitive-ELISA as the method for the quantitative detection. It can detect Trimethoprim (TMP) in samples, such as muscle, feed, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, standard liquid and other supplementary reagents. The microtiter plate in this kit has been pre-coated with antibodies. During the reaction, TMP in the samples or standard competes with Horseradish Peroxidase (HRP) conjugate for sites. Add substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of TMP. The concentration of TMP in the samples can be calculated by comparing the OD of the samples to the standard curve.

Technical indicator

Reaction mode (Incubation time and temperature): 37°C; 45 min, 15 min

Detection limit: Feed---1.6 ppb; Muscle, Kidney, Liver ---0.4 ppb; Serum, Urine ---0.4 ppb;
Egg ---0.3 ppb.

Cross-reactivity: Trimethoprim---100%, Sulfapyridine, Sulfamethoxazole, Sulfisoxazole,
Sulfathiazole, Sulfamerazine, Sulfadoxine ---<0.1%

Sample recovery rate: Feed---95% ± 15%; Muscle, Liver, Kidney---95% ± 10%;
Serum, Urine, Honey ---85% ± 10%

Kits components

Item	Specifications
ELISA Microtiter plate	96 wells
Standard Liquid	1 mL each (ppb=ng/mL=ng/g) (0 ppb, 0.03 ppb, 0.09 ppb, 0.27 ppb, 0.81 ppb, 2.43 ppb)
HRP Conjugate	5.5 mL
Substrate Reagent A	6 mL
Substrate Reagent B	6 mL
Stop Solution	6 mL
20×Concentrated Wash Buffer	40 mL
2×Reconstitution Buffer	50 mL
Plate Sealer	3 pieces
Sealed Bag	1 piece
Manual	1 copy

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

Other materials required but not supplied

Instruments: Microplate reader, Printer, Homogenizer, Nitrogen evaporators, Water bath, Vortex mixer, Centrifuge, Graduated pipette, Balance (sensitivity 0.01 g).

Micropipette: Single channel (20-200 μL , 100-1000 μL), Multichannel (30-300 μL).

Reagents: Methanol anhydrous, N-hexane, HCl, NaOH, Acetonitrile.

Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. FOR RESEARCH USE ONLY. ELISA Microtiter plate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the E-FS-E020. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other E-FS-E020 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue color. When OD value of standard (concentration: 0) <0.5 unit (A450nm <0.5), it indicates the reagent be deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. **For mentioned sample fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

Storage and expiry date

Store the kit at 2-8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2-8°C.

Expiry date: expiration date is on the packing box.

Experimental preparation.

Restore all reagents and samples to room temperature before use.

Open the micro-plate reader in advance, preheat the instrument, and set the testing parameters.

1. Sample pretreatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid cross-contamination during the experiment.

2. Solution preparation

Please prepare solution according to the number of samples. Don't use up all components in the kit at once!

Solution 1: Reconstitution Buffer

Dilute **2×Reconstitution Buffer** with deionized water (**2×Reconstitution Buffer (V): Deionized water (V) = 1:1**). The Reconstitution buffer can be store at 4°C for a month.

Solution 2: 0.1 M HCl Solution (*for feed sample*)

Dilute 10 mL of **HCl** to 1200 mL with deionized water, mix fully.

Solution 3: 1 M NaOH Solution (*for feed sample*)

Dissolve 4 g of **NaOH** to 100 mL with deionized water.

Solution 4: Wash Buffer

Dilute **20×Concentrated Wash Buffer** with deionized water (**20×Concentrated Wash Buffer (V): Deionized water (V) = 1:19**).

3. Sample pretreatment procedure

3.1 Pretreatment of feed sample:

- (1) Homogenize the representative sample with a homogenizer and mix fully.
- (2) Weigh 2 ± 0.05 g of homogenate sample into a 50 mL centrifugal tube, add 20 mL of **0.1 M HCl Solution** (Solution 2) and vortex for 15 min. Centrifuge at 3000 rpm at room temperature for 10 min.
- (3) Take 1 mL of the supernatant into 1.5 mL centrifugal tube, adjust the pH to 6-8 with **1 M NaOH Solution** (Solution 3) (The added amount of 1 M NaOH Solution is different according to the feed sample. The needed amount is generally between 70 μ L-100 μ L), centrifuge at 3000 rpm at room temperature for 10 min.
- (4) Take 0.5 mL of the supernatant into another 1.5 mL centrifugal tube, add 0.5 mL of **Reconstitution Buffer** (Solution 1), and mix thoroughly.
- (5) Take 50 μ L for analysis.

Note: Sample dilution factor: 20, detection limit: 1.6 ppb

3.2 Pretreatment of muscle, liver, kidney sample:

- (1) Remove fat from sample. Homogenize the representative sample with a homogenizer and mix fully.
- (2) Weigh 2 ± 0.05 g of homogenate sample into a 50mL centrifugal tube, add 6 mL of **Methanol anhydrous** and 2 mL of **N-hexane**, vortex for 5 min.
- (3) Centrifuge at 4000 rpm at room temperature for 10 min, remove the upper N-hexane layer, take 0.5 mL of lower liquid into clean glass test tube (do not touch the fat layer) and dry at 50-60°C with nitrogen evaporators or water bath.
- (4) Add 400 μ L of **Reconstitution Buffer** (Solution 1) and 500 μ L of **N-hexane**, vortex for 1 min. Centrifuge at 4000 rpm at room temperature for 5 min.
- (5) Remove upper N-hexane layer, take 50 μ L of lower liquid for analysis.

Note: Sample dilution factor: 5, detection limit: 0.4 ppb

3.3 Pretreatment for urine, serum (swine) sample:

- (1) Weigh 0.5 mL of sample, centrifuge at 4000 rpm at room temperature for 5 min.
- (2) Take 50 μ L of supernatant, add 200 μ L of **Reconstitution Buffer** (Solution 1), and mix thoroughly.
- (3) Take 50 μ L for analysis.

Note: Sample dilution factor: 5, detection limit: 0.4 ppb

3.4 Pretreatment for egg sample:

- (1) Use a homogenizer to homogenize the yolk and egg whites and mix them thoroughly.
- (2) Weigh 2 ± 0.05 g of homogenate sample into a 50 mL centrifugal tube, add 8 mL of **Acetonitrile** and vortex for 15 min. Centrifuge at 4000 rpm at room temperature for 10 min.
- (3) Take 1 mL of supernatant into clean glass test tube (do not touch the fat layer) and dry at 50-60°C with nitrogen evaporators or water bath.
- (4) Dissolve the residue with 1 mL of **N-hexane**, and add 1 mL of **Reconstitution Buffer** (Solution 1), and mix thoroughly. Centrifuge at 4000 rpm at room temperature for 5 min.
- (5) Take 50 μ L for analysis.

Note: Sample dilution factor: 4, detection limit: 0.3 ppb

Assay procedure

Restore all reagents and samples to room temperature (25°C) before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2-8°C.

1. **Number:** Number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples need test in duplicate.**
2. **Add Sample:** add 50 µL of **Standard or Sample** to each well, then add 50 µL of **HRP Conjugate** to each well, cover the plate with plate sealer, oscillate for 5s gently to mix thoroughly, incubate at 37°C for 45 min in shading light.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 300 µL of **Wash Buffer** (Solution 4) to each well and wash. Repeat wash procedure for 5 times, 30s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Color Development:** add 50 µL of **Substrate Reagent A** to each well, and then add 50 µL of **Substrate Reagent B**. Gently oscillate for 5s to mix thoroughly. Incubate at 37°C for 15 min in shading light. (If the blue is too shallow, the reaction time can be prolonged appropriately).
5. **Stop Reaction:** add 50 µL of **Stop Solution** to each well, oscillate gently to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 10 min after stop reaction.

Result analysis

1. Absorbance (%) = $A/A_0 \times 100\%$

A: Average absorbance of standard or sample

A_0 : Average absorbance of 0 ppb Standard

2. Drawing and calculation of standard curve

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis on a large number of samples.

