### AFB1 (Aflatoxin B1) Lateral Flow Assay Kit

Catalog No: E-TO-C001 50T

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017 Email: <u>techsupport@elabscience.com</u> Website: <u>www.elabscience.com</u>

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

## **Test principle**

This kit uses the principle of Immunochromatography assay for the qualitative detection. It can detect Aflatoxin B1 (AFB1) in samples, such as grain, feed, etc. After adding the sample solution into the sample well of detection card, AFB1 in the sample solution combine with the gold-labelled antibody, so as to prevent the combining of gold-labelled antibody with AFB1 conjugate on the cellulose membrane. When the concentration of AFB1 in the sample solution is more than the detection limit, the detect line do not show color and the result is positive. When the concentration of AFB1 in the sample solution is negative.

## **Technical indicator**

**Detection limit:** Grain, Feed ---20 ppb; Oil ---10 ppb Grain, Feed (need to be dry) --- 5 ppb

## **Kits components**

Item	Specifications		
Detection card (with pipette)	50 T/kit		
Manual	1 copy		

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

## Other materials required but not supplied

Instruments: Homogenizer, Oscillators, Centrifuge, Graduated pipette, Balance (sensibility 0.01 g), Nitrogen Evaporators, Water bath.

High-precision transferpettor: Single channel (20-200 µL, 100-1000 µL).

Reagents: Methanol, N-hexane.

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- 2. The detection card should be adjusted to room temperature after removed from the refrigerator before opening. The opening detection card should be used as soon as possible so as not to be invalid because of moisture.
- 3. Avoid of contacting the white membrane at the middle of the sample well.
- 4. The pipette cannot be mixing to avoid the cross-contaminant.
- 5. The tested sample should be clear, no turbidity particle and no bacterial pollution, otherwise it is easy to result in abnormal phenomena such as obstruction, unobvious color, etc., which affect the judgment of the experiment result.
- 6. If the samples are not indicated in the manual, a preliminary experiment to determine the validity of the kit is necessary.
- 7. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.
- 8. Each reagent is optimized for use in the E-TO-C001. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other E-TO-C001 with different lot numbers.

## Storage and expiry date

**Storage:** Store at 2-30°C. With cool and dry environment. **Expiry date:** expiration date is on the packing box.

# Sample pretreatment

Restore all reagents and samples to room temperature before use.

# 1. Sample pretreatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid the experiment result be interfered by the contamination.

# 2. Reagent preparation

Solution 1: 70% Methanol Methanol (V): deionized water (V) =7:3.

#### 3. Sample pretreatment procedure:

### 3.1 Pretreatment of grain, feed sample:

(1) Weigh  $2\pm0.05$  g of crushed homogenate, add **70% Methanol** according to the different detection limit as the following table:

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Detection limit	20 ppb	50 ppb	100 ppb
70% Methanol	4 mL	10 mL	20 mL

Oscillate for 5 min. Centrifuge at 4000 r/min for 5 min at room temperature.

(2) Take 0.1 mL of the supernatant, add 0.4 mL of deionized water. Mix thoroughly to be used. Note: Detection limit: 20 ppb

### 3.2 Pretreatment of grain, feed (need to be dry) sample:

If the detection limit is 5 ppb, you can operate as follows:

- (1) Weigh 2±0.05 g of crushed homogenate, add 4 mL of 70% Methanol. Oscillate hardly for 5 min and centrifuge at 4000 r/min for 5 min at room temperature.
- (2) Take 1 mL of the supernatant, dry the liquid at  $50-60^{\circ}$ C with nitrogen evaporators or water bath.
- (3) Add 0.25 mL of **70% Methanol** to dissolve the rest of the residue, oscillate for 2 min. Add 1 mL of deionized water, mix thoroughly to be used. (Note: the order of 70% Methanol and water cannot be changed at this step.)

### Note: Detection limit: 5 ppb

### 3.3 Pretreatment of oil (vegetable oil, sesame oil, salad oil, peanut oil) sample:

(1) Weigh 2±0.05 g of sample, add **70% Methanol** according to the different detection limit as the following table:

Detection limit	10 ppb	20 ppb	40 ppb	100 ppb
70% Methanol	2 mL	4 mL	8 mL	20 mL

Add 8 mL of N-hexane. Oscillate for 5 min and centrifuge at 4000 r/min for 5 min at room temperature.

(2) Discard the supernatant and take 0.1 mL of the lower layer liquid. Add 0.4 mL of deionized water, mix thoroughly for use.

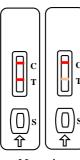
### Note: Detection limit: 10 ppb

### **Experiment** procedure

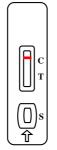
- Tear the aluminum foil bag of the detection card and take out the detection card, and put it on a 1. smooth. clean table.
- 2. Take the prepared clear sample supernatant with the matching pipette, add 2-3 drops (about 60  $\mu$ L) of sample to the sample well (S) vertically and slowly (Avoid foaming).
- 3. Incubate for 8 to 10 minutes and then judge the results immediately.

### Judgment of result

- 1. **Negative:** The test line region (T) and the control line region (C) show a line at the same time in the observation well. It indicates the content of AFB1 in the sample is lower than detection limit or the sample doesn't contain AFB1.
- 2. **Positive:** Only the control line region (C) show a line in the observation well. It indicates the content of AFB1 in the sample is higher than detection limit.
- 3. **Invalid:** The control line region (C) does not show a line in the observation well. It indicates operation process is wrong or the test card is invalid.



Negative



Positive

Invalid