Protocol for Thawing Frozen Cells

The following protocol describes a general procedure for the thawing of cryopreserved cells. For detailed protocols, always refer to the cell-specific product insert.

1) Take the cryovial containing the frozen cells from liquid nitrogen and immediately place it into a 37°C water bath.
2) Quickly thaw the cells (< 1 minute) by gently swirling the vial in the 37°C water bath until there is just a small bit of ice left in the vial.
3) Wipe the outside of the vial with 70% ethanol and transfer the vial into a laminar flow hood.
4) Transfer the appropriate amount of pre-warmed complete growth medium and the thawed cells into a centrifuge tube.
5) Centrifuge the cell suspension at approximately 500 × g for 5-10 minutes. **The actual centrifugation speed and duration varies depending on the cell type.**
6) Discard the supernatant and keep the cell pellet.
7) Gently resuspend the cells with complete growth medium, and transfer into the appropriate culture vessel and into the recommended culture environment.

[Note]:

The actual centrifugation speed and duration varies depending on the cell type.

The appropriate flask size depends on the number of cells frozen in the cryovial.

The culture environment varies based on the cell and medium type.