

(FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !)

Catalog No: E-BC-K171-S

Specification: 50 assays(36 samples)/100 assays(86 samples)

Measuring instrument: Spectrophotometer (370 nm)

Detection range: 0.94-45 µg/mL

Elabsience[®] Total Carbonyl Colorimetric Assay Kit

This manual must be read attentively and completely before using this product.

If you have any problem, please contact our Technical Service Center for help:

Toll-free: 1-888-852-8623

Tell: 1-832-243-6086

Fax: 1-832-243-6017

Email: techsupport@elabsience.com

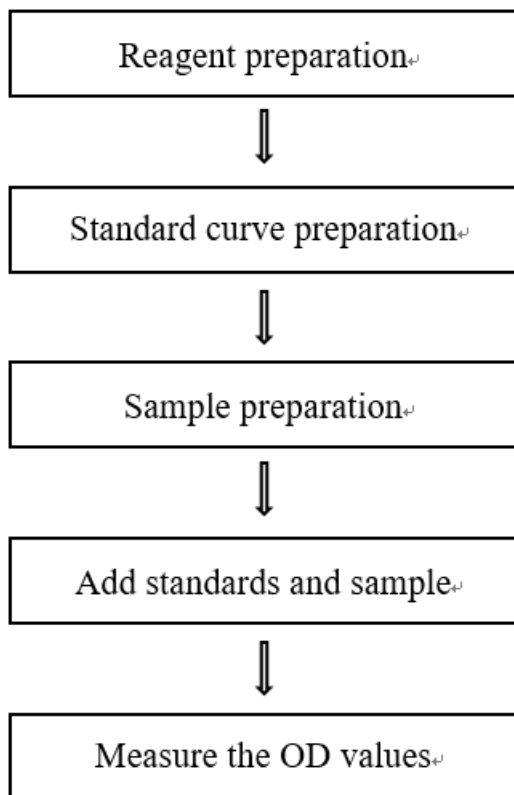
Website: www.elabsience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

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Assay summary

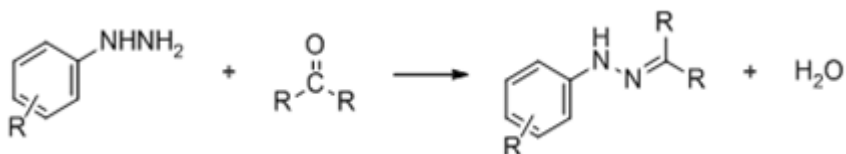


Intended use

This kit can be used for detection of total carbonyl content in serum, plasma and tissue samples.

Detection principle

Carbonyl can react with 2,4-dinitrophenylhydrazine and produce a kind of reddish brown hydrazone compounds, which has a specific absorbance peak at 370 nm. The content of carbonyl can be calculated according to the absorbance value.



Kit components & storage

Item	Component	Size 1 (50 assays)	Size 2 (100 assays)	Storage
Reagent 1	Working Solution	15 mL × 1 vial	30 mL × 1 vial	2-8 °C, 12 months, shading light
Reagent 2	100 µg/mL Standard	1 mL × 1 vial	2 mL × 1 vial	2-8 °C, 12 months

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

Materials prepared by users

Instruments:

Spectrophotometer (370 nm), Micropipettor, Vortex mixer.

Reagents:

Double distilled water, Normal saline (0.9% NaCl), PBS (0.01 M, pH 7.4)

Reagent preparation

① Equilibrate all the reagents to room temperature before use.

② The preparation of standard curve:

Always prepare a fresh set of standards. Discard working standard dilutions after use.

Dilute 100 $\mu\text{g}/\text{mL}$ standard with double distilled water diluent to a serial concentration. The recommended dilution gradient is as follows: 5, 10, 20, 30, 40, 45 $\mu\text{g}/\text{mL}$. Reference is as follows:

Item	①	②	③	④	⑤	⑥
Concentration ($\mu\text{g}/\text{mL}$)	5	10	20	30	40	45
100 $\mu\text{g}/\text{mL}$ standard (μL)	13	26	52	78	104	117
Double distilled water (μL)	247	234	208	182	156	143

Sample preparation

① Sample preparation:

Serum and plasma: detect directly. If not detected on the same day, the serum or plasma can be stored at -80 °C for a month.

Tissue samples:

- ① Harvest the amount of tissue needed for each assay (initial recommendation 20 mg).
- ② Wash tissue in cold PBS (0.01 M, pH 7.4).
- ③ Homogenize 20 mg tissue in 180 μ L PBS (0.01 M, pH 7.4) with a dounce homogenizer at 4 °C.
- ④ Centrifuge at 10000 \times g for 10 min to remove insoluble material. Collect supernatant and keep it on ice for detection.

② Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Total carbonyl content (μ g/mL)	Dilution factor
< 45	1
45-450	10

Note: The diluent is normal saline (0.9% NaCl) or PBS (0.01 M, pH 7.4). For the dilution of other sample types, please do pretest to confirm the dilution factor.

The key points of the assay

The supernatant of sample must be clarified.

Operating steps

- ① Blank tube: add 1.62 mL of double distilled water to the 2 mL EP tube.
Standard tube: add 1.5 mL of double distilled water and 0.12 mL of standard with different concentrations to the 2 mL EP tube.
Sample tube: add 1.5 mL of double distilled water and 0.12 mL of sample to the 2 mL EP tube.
- ② Add 0.25 mL of working solution and oscillate fully.
- ③ Stand for 5 min at room temperature. Set the spectrometer to zero with double distilled water and measure the OD values of each tube at 370 nm with 0.5 cm optical path cuvette.

Calculation

The standard curve:

1. Average the duplicate reading for each standard.
2. Subtract the mean OD value of the blank (Standard #①) from all standard readings. This is the absolved OD value.
3. Plot the standard curve by using absolved OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve ($y = ax + b$) with graph software (or EXCEL).

The sample:

1. Serum (plasma) sample:

$$\begin{array}{l} \text{Total carbonyl content} \\ (\mu\text{g/mL}) \end{array} = (\Delta A_{370} - b) \div a \times f$$

2. Tissue sample:

$$\begin{array}{l} \text{Total carbonyl content} \\ (\mu\text{g/g}) \end{array} = (\Delta A_{370} - b) \div a \div c \times f$$

[Note]

ΔA_{370} : Absolute OD ($OD_{\text{Sample}} - OD_{\text{Blank}}$).

f: Dilution factor of sample before test.

c: The content of sample = the wet weight (g) \div the volume of homogenized medium (mL).

Appendix I Performance Characteristics

1. Parameter:

Intra-assay Precision

Three human serum samples were assayed in replicates of 20 to determine precision within an assay. (CV = Coefficient of Variation)

Parameters	Sample 1	Sample 2	Sample 3
Mean ($\mu\text{g/mL}$)	4.60	18.50	36.20
%CV	4.7	4.2	4.3

Inter-assay Precision

Three human serum samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3
Mean ($\mu\text{g/mL}$)	4.60	18.50	36.20
%CV	8.3	8.1	8.5

Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 100%.

	Standard 1	Standard 2	Standard 3
Expected Conc. ($\mu\text{g/mL}$)	12.5	26.8	39.2
Observed Conc. ($\mu\text{g/mL}$)	12.6	28.1	36.8
Recovery rate (%)	101	105	94

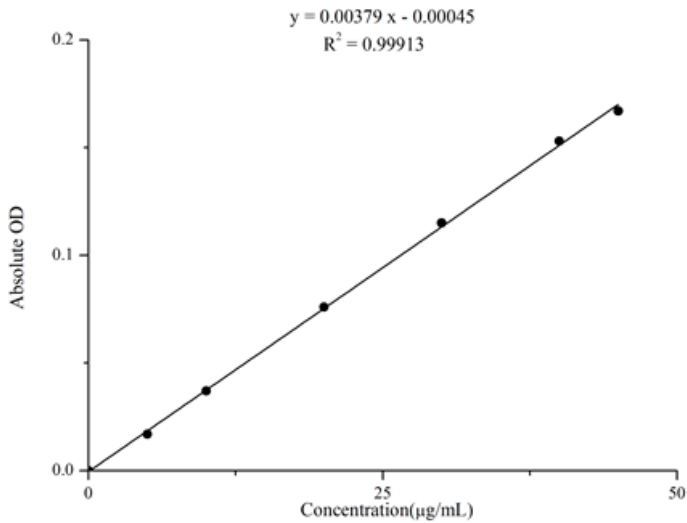
Sensitivity

The analytical sensitivity of the assay is $0.94 \mu\text{g/mL}$. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

2. Standard curve

As the OD value of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique or temperature effects), so the standard curve and data are provided as below for reference only.

Concentration ($\mu\text{g/mL}$)	0	5	10	20	30	40	45
Average OD	0.358	0.375	0.395	0.434	0.473	0.511	0.525
Absoluted OD	0	0.017	0.037	0.076	0.115	0.153	0.167



Appendix II Example Analysis

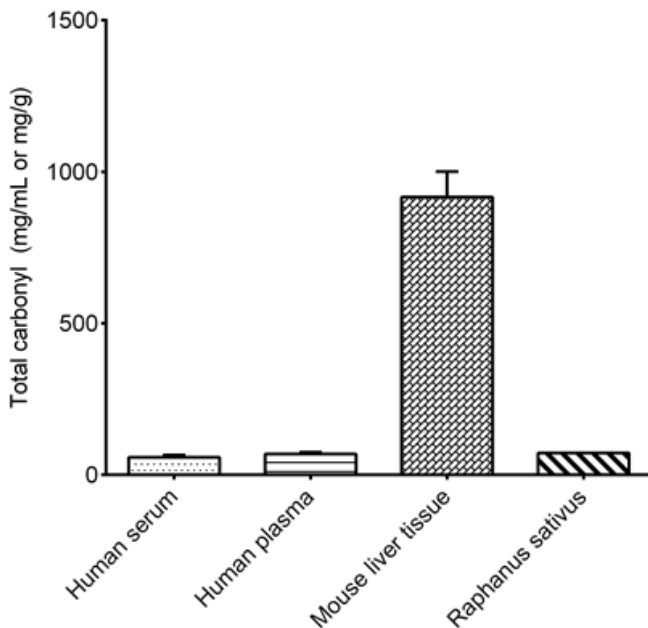
Example analysis:

Take 10% mouse liver tissue homogenate, then dilute the supernatant with PBS for 6 times, take 0.12 mL of diluted sample, and carry the assay according to the operation steps. The results are as follows:

Standard curve: $y = 0.00366x + 0.00354$, $R^2 = 0.99719$. The average OD value of the sample is 0.445, the average OD value of the blank is 0.379, and the calculation result is:

$$\text{Total carbonyl content} \begin{matrix} (\mu\text{g/g}) \end{matrix} = (0.445 - 0.379 - 0.00354) \div 0.00366 \div (0.1 \text{ g} \div 0.9 \text{ mL}) \times 6 = 917 \mu\text{g/g}$$

Detect human serum, human plasma, mouse liver tissue (the concentration of protein is 0.111 gprot/mL), Raphanus sativus (the concentration of protein is 0.111 gprot/mL) according to the protocol, the result is as follows:



Statement

1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
3. Protection methods must be taken by wearing lab coat and latex gloves.
4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.