#### (FOR RESEARCH USE ONLY, DO NOT USE IT IN CLINICAL DIAGNOSIS!)

Catalog No: E-BC-K751-M

**Specification:** 96T(40 samples)

Measuring instrument: Microplate reader (500-520 nm)

Detection range: 20-2000 U/mL

# Elabscience® Sucrase Activity Assay Kit

This manual must be read attentively and completely before using this product. If you have any problem, please contact our Technical Service Center for help:

Toll-free: 1-888-852-8623

Tell: 1-832-243-6086

Fax: 1-832-243-6017

Email: tech support@elabscience.com

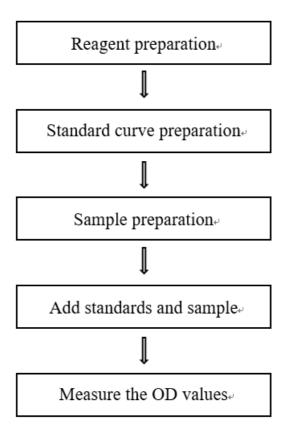
Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

# **Table of contents**

| Assay summary                          | 3  |
|--|----|
| Intended use                           | 4  |
| Detection principle                    | 4  |
| Kit components & storage               | 4  |
| Materials prepared by users            | 5  |
| Reagent preparation                    | 5  |
| Sample preparation                     | 6  |
| The key points of the assay            | 7  |
| Operating steps                        | 7  |
| Calculation                            | 8  |
| Appendix I Performance Characteristics | 9  |
| Appendix Π Example Analysis            | 11 |
| Statement                              | 12 |

# **Assay summary**



#### Intended use

This kit can be used to measure sucrase activity in animal tissue samples.

# **Detection principle**

Sucrase catalyzes its substrate (sucrose) to produce glucose, which produces hydrogen peroxide under the action of glucose oxidase. Hydrogen peroxide reacts with chromogenic agent to produce red substance, which has a strong absorption peak at 505 nm. In a certain concentration range, It's absorbance is proportional to glucose concentration. Therefore, the activity of sucrase can be calculated by measuring the OD value at 505 nm.

### **Kit components & storage**

| Item                                 | Component       | Size (96 T)    | Storage                            |
|--------------------------------------|-----------------|----------------|------------------------------------|
| Reagent 1                            | Substrate       | Powder ×1 vial | 2-8°C, 12 months                   |
| Reagent 2                            | Buffer Solution | 10 mL ×1 vial  | 2-8°C, 12 months                   |
| Reagent 3                            | Phenol Solution | 12 mL ×1 vial  | 2-8°C, 12 months,<br>shading light |
| Reagent 4                            | Enzyme Solution | 12 mL ×1 vial  | 2-8°C, 12 months,<br>shading light |
| Reagent 5                            | Stop Solution   | Powder ×1 vial | 2-8°C, 12 months                   |
| Reagent 6 50 mmol/L Glucose Standard |                 | 1 mL ×1 vial   | 2-8°C, 12 months                   |
|                                      | Microplate      | 96 wells       | No requirement                     |
|                                      | Plate Sealer    | 2 pieces       |                                    |

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

# Materials prepared by users

#### **Instruments:**

Micropipette, Vortex mixer, Centrifuge, Microplate reader (500-520 nm, optimum wavelength: 505 nm)

#### **Reagents:**

Double distilled water, PBS (0.01 M, pH 7.4)

# Reagent preparation

- ① Equilibrate all reagents to room temperature before use.
- ② Preparation of substrate working solution:

  Dissolve one vial of substrate with 8 mL of buffer solution, mix well to dissolve. Store at 2-8 ℃ for 7 days.
- ③ Preparation of stop working solution:

  Dissolve one vial of stop solution with 5 mL of ultrapure water, mix well to dissolve. Store at 2-8 ℃ for 7 days.
- 4 Preparation of chromogenic solution: For each well, prepare 200  $\mu$ L of chromogenic solution (mix well 100  $\mu$ L of phenol solution and 100  $\mu$ L of enzyme solution). The chromogenic solution should be prepared on spot.
- (5) The preparation of standard curve:

  Always prepare a fresh set of standards. Discard working standard dilutions after use.

Dilute 50 mmol/L standard solution with double distilled water to a serial concentration. The recommended dilution gradient is as follows: 0, 1, 2, 5, 10, 15, 20, 25 mmol/L. Reference is as follows:

| Item                        | 1 | 2   | 3   | 4   | (5) | 6   | 7   | 8   |
|-----------------------------|---|-----|-----|-----|-----|-----|-----|-----|
| Concentration (mmol/L)      |   | 1   | 2   | 5   | 10  | 15  | 20  | 25  |
| 50 mmol/L standard (μL)     |   | 10  | 20  | 50  | 100 | 150 | 200 | 250 |
| Double distilled water (μL) |   | 490 | 480 | 450 | 400 | 350 | 300 | 250 |

# Sample preparation

## **1** Sample preparation

## Tissue sample:

- Harvest the amount of tissue needed for each assay (initial recommendation 20 mg).
- ② Wash tissue in cold PBS (0.01 M, pH 7.4).
- 3 Homogenize 20 mg tissue in 80  $\mu$ L PBS (0.01 M, pH 7.4) with a dounce homogenizer at 4°C.
- ④ Centrifuge at 10000 xg for 10 min at 4°C to remove insoluble material. Collect supernatant and keep it on ice for detection.
- (E-BC-K318-M).

### 2 Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

| Sample type                       | Dilution factor |
|-----------------------------------|-----------------|
| 20% Rat ileum tissue homogenate   | 1               |
| 20% Rat stomach tissue homogenate | 1               |
| 20% Rat liver tissue homogenate   | 1               |

Note: The diluent is PBS (0.01 M, pH 7.4). For the dilution of other sample types, please do pretest to confirm the dilution factor.

# The key points of the assay

- ① Control the time of enzymatic reaction strictly.
- ② Avoid contaminating enzyme solution when preparing of chromogenic solution.

# **Operating steps**

① Standard tube: Take 25  $\mu$ L of standards with different concentrations into 1.5 mL EP tubes.

Sample tube: Take 25  $\mu$ L of sample into 1.5 mL EP tubes.

Control tube: Take 50 µL of substrate working solution into 1.5 mL EP tubes.

- $\odot$  Add 50  $\mu$ L of substrate working solution into the standard tubes and sample tubes.
- ③ Mix fully and incubate at 37°C for 20 min.
- 4 Add 25  $\mu$ L of stop working solution into each tube and mix fully. Add 25  $\mu$ L of sample into control tubes.
- $^{\circ}$  Mix fully, centrifuge at 1780×g for 10 min and take 8  $\mu$ L of supernatant from each tube to the corresponding wells.
- ⑥ Add 200 μL of chromogenic solution into each well.
- (7) Mix fully with microplate reader for 10 s, incubate at 37°C for 15 min. Measure the OD values of each well at 505 nm with microplate reader.

#### **Calculation**

#### The standard curve:

1. Average the duplicate reading for each standard.

2. Subtract the mean OD value of the blank (Standard # 1) from all standard

readings. This is the absoluted OD value.

3. Plot the standard curve by using absoluted OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve (y = ax + b) with graph software (or EXCEL).

## The sample:

# **Tissue sample:**

**Definition:** The amount of 1 nmol sucrose hydrolysised by 1 g tissue protein per minute at 37°C is defined as 1 activity unit

Sucrase activity (U/mgprot) =  $(\Delta A - b) \div a \div T \times 1000^* \times f \div C_{pr}$ 

### [Note]

 $\Delta A$ :  $OD_{Sample} - OD_{Control}$ .

T: Enzymatic reaction reaction time, 20 min.

 $1000*: 1 \mu mol = 1000 nmol.$ 

f: Dilution factor of sample before test.

 $C_{\text{pr}}$ : Concentration of protein in tissue sample, mgprot/mL.

# **Appendix I Performance Characteristics**

#### 1. Parameter:

#### **Intra-assay Precision**

Three rat liver tissue samples were assayed in replicates of 20 to determine precision within an assay (CV = Coefficient of Variation).

| Parameters  | Sample 1 | Sample 2 | Sample 3 |  |  |
|-------------|----------|----------|----------|--|--|
| Mean (U/mL) | 32.50    | 135.00   | 482.00   |  |  |
| %CV         | 5.7      | 5.3      | 5.2      |  |  |

#### **Inter-assay Precision**

Three rat liver tissue samples were assayed 20 times in duplicate by three operators to determine precision between assays.

| Parameters  | Sample 1 | Sample 2 | Sample 3 |  |
|-------------|----------|----------|----------|--|
| Mean (U/mL) | 32.50    | 135.00   | 482.00   |  |
| %CV         | 6.2      | 6.4      | 6.9      |  |

## Recovery

Take three samples of high concentration, middle concentration and low concentration to test the samples of each concentration for 6 times parallelly to get the average recovery rate of 100%.

|                         | Standard 1 | Standard 2 | Standard 3 |
|-------------------------|------------|------------|------------|
| Expected Conc. (mmol/L) | 1.5        | 7.8        | 17.5       |
| Observed Conc. (mmol/L) | 1.5        | 8.0        | 16.8       |
| Recovery rate (%)       | 101        | 103        | 96         |

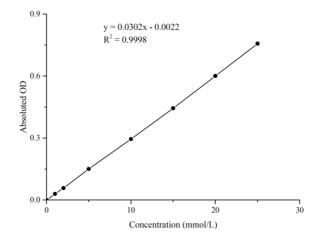
### **Sensitivity**

The analytical sensitivity of the assay is 20 U/mL. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

## 2. Standard curve:

As the OD value of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique or temperature effects), so the standard curve and data are provided as below for reference only:

| Concentration (mmol/L) | 0     | 1     | 2     | 5     | 10    | 15    | 20    | 25    |
|------------------------|-------|-------|-------|-------|-------|-------|-------|-------|
| Average OD             | 0.083 | 0.112 | 0.140 | 0.233 | 0.378 | 0.527 | 0.683 | 0.840 |
| Absoluted OD           | 0.000 | 0.030 | 0.058 | 0.150 | 0.295 | 0.444 | 0.600 | 0.757 |



# **Appendix Π Example Analysis**

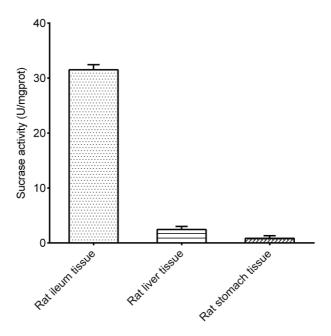
#### **Example analysis:**

For rat ileum tissue, take 25  $\mu$ L of rat ileum tissue supernatant and carry the assay according to the operation table. The results are as follows:

standard curve: y = 0.0306 x + 0.0025, the average OD value of the sample is 0.204, the average OD value of the control is 0.079, the concentration of protein in sample is 6.48 mgprot/mL, and the calculation result is:

Sucrase activity (U/mgprot) = 
$$(0.204-0.079-0.0025 \div 0.0306 \div 20 \times 1000 \div 6.48$$
  
=  $30.89$  U/mgprot

Detect 20% rat ileum tissue homogenate (the concentration of protein is 6.48 mgprot/mL), 20% rat liver tissue homogenate (the concentration of protein is 23.65 mgprot/mL) and 20% rat stomach tissue homogenate (the concentration of protein is 5.89 mgprot/mL) according to the protocol, the result is as follows:



#### Statement

- 1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
- 2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
- 3. Protection methods must be taken by wearing lab coat and latex gloves.
- 4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
- 5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
- 6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.