# Elabscience<sup>®</sup>Lactate Dehydrogenase (LDH) Activity Assay Kit (WST-8 Method)

Catalog No: E-BC-K766-M Specification: 48T(32 samples)/96T(80 samples) Measuring instrument: Microplate reader (450 nm) Detection range: 0.11-39.9 U/L

This manual must be read attentively and completely before using this product. If you have any problem, please contact our Technical Service Center for help:

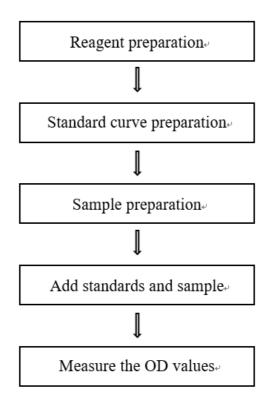
Toll-free: 1-888-852-8623 Tell: 1-832-243-6086 Fax: 1-832-243-6017 Email: techsupport@elabscience.com Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

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## Intended use

This kit can be used to measure lactate dehydrogenase (LDH) activity in tissues, serum (plasma), hydrothorax, cells and cell culture supernatant samples.

# **Detection principle**

Lactate dehydrogenase catalyzes the reaction of lactic acid with NAD<sup>+</sup> to produce pyruvic acid and NADH. NADH, under the action of PMS, transfer electrons to WST-8 to produce the yellow product, which has a characteristic absorption peak at 450 nm.

Item	Component	Size 1(48 T)	Size 2(96 T)	Storage
Reagent 1	Lysis Solution	$60 \text{ mL} \times 1 \text{ vial}$	$60 \text{ mL} \times 2 \text{ vials}$	-20°C, 12 months
Reagent 2	Substrate	$1.5 \text{ mL} \times 1 \text{ vial}$	1.5 mL $\times$ 2 vials	-20°C, 12 months
Reagent 3	Chromogenic Agent	$1.5 \text{ mL} \times 1 \text{ vial}$	$1.5 \text{ mL} \times 2 \text{ vials}$	-20°C, 12 months, shading light
Reagent 4	Coenzyme	Powder $\times 1$ vial	Powder $\times 1$ vial	-20°C, 12 months
Reagent 5	Stop Solution	$3 \text{ mL} \times 1 \text{ vial}$	6 mL ×1 vial	-20°C, 12 months
Reagent 6	NADH Standard	Powder ×1 vial	Powder ×1 vial	-20°C, 12 months
	Microplate	96 wells		No requirement
	Plate Sealer	2 pieces		

## Kit components & storage

Note: The reagents must be stored strictly according to the preservation conditions in the above table. The reagents in different kits cannot be mixed with each other. For a small volume of reagents, please centrifuge before use, so as not to obtain sufficient amount of reagents.

# Materials prepared by users

# Instruments:

Micropipettor, Vortex mixer, Incubator, Centrifuge, Microplate reader (450 nm)

# **Reagents:**

Double distilled water, PBS (0.01 M, pH 7.4)

# **Reagent preparation**

- Preheat stop solution at 37 ℃ for 20 min in advance and can be used only after it is completely clarified. Equilibrate other reagents to room temperature before use.
- ② Preparation of coenzyme working solution: Dissolve one vial of coenzyme with 260 μL of double distilled water, mix well to dissolve. The coenzyme working solution should be prepared on spot. Store at -20 ℃ for 7 days protected from light.
- ③ Preparation of reaction working solution: For each well, prepare 50 µL of reaction working solution (mix well 24 µL of substrate, 24 µL of chromogenic agent and 2 µL of coenzyme working solution). The reaction working solution should be prepared on spot and stored protected from light.
- ④ Preparation of 5 mmol/L standard solution:
  Dissolve one vial of NADH standard with 2 mL of double distilled water, mix well to dissolve. The 5 mmol/L standard solution should be prepared on spot. Store at -20 °C for 7 days protected from light
- 5 The preparation of standard curve:

Always prepare a fresh set of standards. Discard working standard dilutions after use.

Dilute 5 mmol/L NADH standard with lysis solution to a serial concentration. The recommended dilution gradient is as follows: 400, 300, 250, 200, 150,

Item	1	2	3	4	5	6	$\overline{O}$	8
Concentration (µmol/L)	0	50	100	150	200	250	300	400
5 mmol/L NADH standard (µL)	0	20	40	80	100	120	140	160
Lysis solution (µL)	1000	900	980	970	960	950	940	920

100, 50, 0 µmol/L. Reference is as follows:

# **Sample preparation**

## **(1)** Sample preparation

**Serum and plasma:** detect directly. If not detected on the same day, the serum or plasma can be stored at -80°C for a month.

## **Tissue sample:**

- Harvest the amount of tissue needed for each assay (initial recommendation 20 mg).
- ② Wash tissue in cold PBS (0.01 M, pH 7.4).
- 3 Homogenize 20 mg tissue in 180  $\mu$ L lysis solution with a dounce homogenizer at 4°C.
- (4) Centrifuge at 10000×g for 10 minutes to remove insoluble material. Collect supernatant and keep it on ice for detection.
- (5) Meanwhile, determine the protein concentration of supernatant (E-BC-K318-M).

## Cell (adherent or suspension) samples:

- (1) Harvest the number of cells needed for each assay (initial recommendation =  $1 \times 10^{6}$  cells).
- 2 Wash cells with PBS (0.01 M, pH 7.4).
- (3) Lyse  $1 \times 10^{6}$  cells with 400 µL lysis solution. Place on the ice box and mix well every 5 min, lyse for 10 min.
- ④ Centrifuge at 15000×g for 10 min. Collect supernatant and keep it on ice for detection.

⑤ Meanwhile, determine the protein concentration of supernatant

(E-BC-K318-M).

**Cell culture supernatant:** Collect fresh cell culture supernatant and centrifuge at 10000×g for 10 min at 4°C. Take the supernatant to preserve it on ice for detection.

# **②** Dilution of sample

The recommended dilution factor for different samples is as follows (for reference only):

Sample type	Dilution factor
Human serum	10-20
Dog serum	10-20
Mouse serum	50-100
Cynomolgus monkey serum	10-20
10% Rat spleen tissue homogenate	150-250
10% Rat liver tissue homogenate	250-350
10% Rat kidney tissue homogenate	250-350
10% Rat lung tissue homogenate	250-350

Note: The diluent is lysis solution. For the dilution of other sample types, please do pretest to confirm the dilution factor.

# The key points of the assay

There should be no bubbles in the wells of the microplate when measuring the OD value.

## **Operating steps**

(1) Standard well: Take 50  $\mu$ L of standard solution with different concentrations into the corresponding wells.

Sample well: Take 50 µL of sample into the corresponding wells.

- (2) Add 50 µL of reaction working solution to each well.
- ③ Incubate at 37°C for 10 min.
- (4) Add 50  $\mu$ L of stop solution to each well.
- (5) Mix fully for 5 s with microplate reader. Measure the OD values of each well with microplate reader at 450 nm.

## Calculation

#### The standard curve:

1. Average the duplicate reading for each standard.

2. Subtract the mean OD value of the blank (Standard #(1)) from all standard readings. This is the absoluted OD value.

3. Plot the standard curve by using absoluted OD value of standard and correspondent concentration as y-axis and x-axis respectively. Create the standard curve ( $\mathbf{y} = \mathbf{ax} + \mathbf{b}$ ) with graph software (or EXCEL).

#### The sample:

#### 1. Serum (plasma), cell culture supernatant and other liquid sample:

**Unit definition:** the enzyme amount of 1  $\mu$ mol of NADH generated by 1 L of liquid sample per minute at 37°C is defined as 1 unit.

LDH activity (U/L) = ( $\Delta A_{450}$  - b)  $\div a \div T \times f$ 

#### 2. Tissue and cells sample:

**Unit definition:** the enzyme amount of 1 µmol of NADH generated by 1 g tissue protein or cell protein per minute at 37°C is defined as 1 unit.

LDH activity (U/gprot) =  $(\Delta A_{450} - b) \div a \div T \times f \div C_{pr}$ 

## [Note]

 $\Delta A_{450}$ : OD<sub>Sample</sub> - OD<sub>Blank</sub>.

f: Dilution factor of sample before test.

T: Reaction time (10 min).

C<sub>pr</sub>: Concentration of protein in sample (gprot/L).

# **Appendix I Performance Characteristics**

## 1. Parameter:

#### **Intra-assay Precision**

Three human serum samples were assayed in replicates of 20 to determine precision

Parameters	Sample 1	Sample 2	Sample 3	
Mean (U/L)	2.60	18.50	26.00	
%CV	2.6	2.3	2.0	

#### **Inter-assay Precision**

Three human serum samples were assayed 20 times in duplicate by three operators to determine precision between assays.

Parameters	Sample 1	Sample 2	Sample 3	
<b>Mean (U/L)</b> 2.60		18.50	26.00	
%CV	2.4	2.5	2.0	

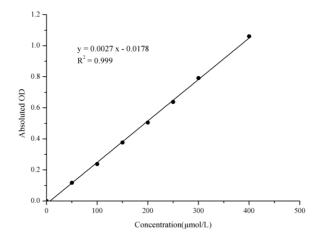
## Sensitivity

The analytical sensitivity of the assay is 0.11 U/L. This was determined by adding two standard deviations to the mean O.D. obtained when the zero standard was assayed 20 times, and calculating the corresponding concentration.

# 2. Standard curve:

As the OD value of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique or temperature effects), so the standard curve and data are provided as below for reference only:

Concentration (µmol/L)	0	50	100	150	200	250	300	400
Average OD	0.054	0.171	0.292	0.430	0.559	0.691	0.845	1.114
Absoluted OD	0.000	0.117	0.238	0.377	0.505	0.638	0.792	1.061



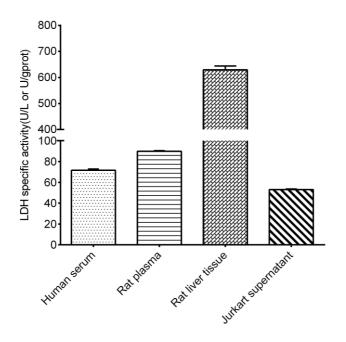
# Appendix II Example Analysis

#### Example analysis:

Take 50  $\mu$ L of human serum diluted for 10 times, and carry the assay according to the operation table. The results are as follows:

Standard curve: y = 0.0027 x - 0.0178, the average OD value of the sample is 0.229, the average OD value of the blank is 0.054, and the calculation result is:

LDH activity (U/L) = (0.229 - 0.054 + 0.0178)  $\div 0.0027 \div 10 \times 10 = 71.4$  U/L Detect human serum (dilute for 10 times), rat plasma (dilute for 10 times), 10% rat kidney tissue homogenate (the concentration of protein is 14.62 gprot/L dilute for 250 times) and Jurkart supernatant (dilute for 3 times) according to the protocol, the result is as follows:



## Statement

- 1. This assay kit is for Research Use Only. We will not response for any arising problems or legal responsibilities causing by using the kit for clinical diagnosis or other purpose.
- 2. Please read the instructions carefully and adjust the instruments before the experiments. Please follow the instructions strictly during the experiments.
- 3. Protection methods must be taken by wearing lab coat and latex gloves.
- 4. If the concentration of substance is not within the detection range exactly, an extra dilution or concentration should be taken for the sample.
- 5. It is recommended to take a pre-test if your sample is not listed in the instruction book.
- 6. The experimental results are closely related to the situation of reagents, operations, environment and so on. Elabscience will guarantee the quality of the kits only, and NOT be responsible for the sample consumption caused by using the assay kits. It is better to calculate the possible usage of sample and reserve sufficient samples before use.