

**OFL (Ofloxacin) ELISA Kit**

Catalog No: E-FS-E129

96T/96T\*3

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Toll-free: 1-888-852-8623 Tel: 1-832-243-6086 Fax: 1-832-243-6017

Email: [techsupport@elabscience.com](mailto:techsupport@elabscience.com)

Website: [www.elabscience.com](http://www.elabscience.com)

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

## Test principle

This kit uses Competitive-ELISA as the method for the quantitative detection. It can detect ofloxacin (OFL) in samples, such as honey, muscle, milk, milk powder, egg, etc. This kit is composed of ELISA Microtiter plate, HRP conjugate, antibody working solution, standard and other supplementary reagents. The microtiter plate in this kit has been pre-coated with coupled antigen. During the reaction, OFL in the samples or standard competes with coupled antigen on the solid phase supporter for sites of anti-OFL antibody. Then Horseradish Peroxidase (HRP) conjugate is added to each microtiter plate well, and substrate reagent is added for color development. There is a negative correlation between the OD value of samples and the concentration of OFL. The concentration of OFL in the samples can be calculated by comparing the OD of the samples to the standard curve.

## Technical indicator

**Reaction mode** (Incubation time and temperature): 25°C; 45 min, 15 min

**Detection limit:** Muscle, Honey---0.3 ppb; Milk---3 ppb; Milk powder---7.5 ppb;  
Egg, Serum---4.5 ppb; Urine ---0.75 ppb.

**Cross-reactivity:** Ofloxacin---100%

**Sample recovery rate:** Muscle, honey, Milk, Milk powder, Egg, Serum---85% ± 15%.

## Kits components

Item	Specifications
ELISA Microtiter plate	96 wells
Standard Liquid	1 mL each (ppb=ng/mL=ng/g) (0 ppb, 1.5 ppb, 3 ppb, 6 ppb, 12 ppb, 24 ppb)
HRP Conjugate	5.5 mL
Antibody Working Solution	5.5 mL
Substrate Reagent A	6 mL
Substrate Reagent B	6 mL
Stop Solution	6 mL
5×Reconstitution Buffer	50 mL
20×Concentrated Wash Buffer	40 mL
Plate Sealer	3 pieces
Sealed Bag	1 piece
Manual	1 copy
HRP Conjugate	5.5 mL

Note: All reagent bottle caps must be tightened to prevent evaporation and microbial pollution.

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### Other materials required but not supplied

**Instrument:** Microtiter plate reader, Printer, Homogenizer, Nitrogen Evaporators, Water bath, Vortex mixer, Oscillators, Centrifuge, Graduated pipette, Balance (sensitivity 0.01 g).

**Micropipette:** Single channel (20-200 µL, 100-1000 µL), Multichannel (30-300 µL).

**Reagents:** N-hexane, Anhydrous acetonitrile, Concentrated HCl, Dichloromethane.

### Notes

1. The overall OD value will be lower when reagents have not been brought to room temperature before use or room temperature is below 25°C.
2. If the wells turn dry during the washing procedure, it will lead to bad linear standard curve and poor repeatability. Operate the next step immediately after wash.
3. Mix thoroughly and wash the plate completely. The consistency of wash procedure can strongly affect the reproducibility of this ELISA kit.
4. FOR RESEARCH USE ONLY. ELISA Microtiter plate should be covered by plate sealer. Avoid the kit to strong light.
5. **Each reagent is optimized for use in the E-FS-E129. Do not substitute reagents from any other manufacturer into the test kit. Do not combine reagents from other E-FS-E129 with different lot numbers.**
6. Substrate Reagent should be abandoned if it turns blue color. When OD value of standard (concentration: 0) < 0.5 unit (A450nm < 0.5), it indicates the reagent may be deteriorated.
7. Stop solution is caustic, avoid contact with skin and eyes.
8. As the OD values of the standard curve may vary according to the conditions of the actual assay performance (e.g. operator, pipetting technique, washing technique or temperature effects), the operator should establish a standard curve for each test.
9. Even the same operator might get different results in two separate experiments. In order to get reproducible results, the operation of every step in the assay should be controlled.
10. **For mentioned sample fast and efficient extraction methods are included in the kit description. Please consult technical support for the applicability if other sample need to be tested.**
11. The kit is used for rapid screening of actual samples. If the test result is positive, the instrument method such as HPLC, LC/MS, etc. can be used for quantitative confirmation.

### Storage and expiry date

Store the kit at 2-8°C. Do not freeze any test kit components.

Return any unused microwells to their original foil bag and reseal them together with the desiccant provided and further store at 2-8°C.

**Expiry date:** expiration date is on the packing box.

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## Experimental preparation

Restore all reagents and samples to room temperature before use.

Open the microplate reader in advance, preheat the instrument, and set the testing parameters

### 1. Sample pretreatment Notice:

Experimental apparatus should be clean, and the pipette should be disposable to avoid cross-contamination during the experiment.

### 2. Solution preparation

*Please prepare solution according to the number of samples. Don't use up all components in the kit at once!*

Solution 1: 0.15M HCl Solution

Dissolve 5 mL of **Concentrated HCl** to 400 mL with deionized water, mix fully.

Solution 2: Sample Extraction Solution (*for muscle, honey sample*)

Take 10 ml **0.15M HCl Solution** (solution 1) and 90 ml of **Anhydrous acetonitrile**, mix fully.

Solution 3: Reconstitution Buffer

Dilute **5×Reconstitution Buffer** with deionized water. (5×Reconstitution Buffer (V): Deionized water (V) = 1:4), it could be stored at 4°C for one month.

Solution 4: Wash Buffer

Dilute **20×Concentrated Wash Buffer** with deionized water. (20× Concentrated Wash Buffer (V): Deionized water (V) = 1:19).

### 3. Sample pretreatment procedure

#### 3.1 Pretreatment of muscle (chicken, pork, fish, shrimp) sample:

- (1) Homogenize the representative sample with a homogenizer and mix fully.
- (2) Weigh  $2 \pm 0.05$  g of homogenized sample into 50 mL centrifuge tube, add 8 mL of **Sample Extraction Solution** (Solution 2), oscillate for 5 min, centrifuge at 4000 rpm for 10 min at room temperature.
- (3) Take 2 mL of supernatant to a 10 mL cool glass tube, dry with nitrogen evaporators/water bath at 50-60°C (Please do it in the fume hood).
- (4) Add 1 mL of **N-hexane**, oscillate for 2 min. Add 1 mL of **Reconstitution Buffer** (Solution 3), oscillate for 30 s. Centrifuge at 4000 rpm for 5 min at room temperature.
- (5) Discard the upper N-hexane, take 50  $\mu$ L lower liquid for analysis.

**Note: Sample dilution factor: 2, detection limit: 3 ppb**

**3.2 Pretreatment of honey sample:**

- (1) Weigh  $1 \pm 0.05$  g of honey sample into 50 mL centrifuge tube, and add 6 mL of **Sample Extraction Solution** (Solution 2), oscillate for 5 min and mix fully.
- (2) Add 3 mL of **Reconstitution Buffer** (Solution 3) and 11 mL of **Dichloromethane**, oscillate for 5 min. Centrifuge at 4000 rpm for 5 min at room temperature.
- (3) Remove the upper liquid, take 8 mL lower organic phase into another cool centrifuge tube, dry with nitrogen evaporators/water bath at 50-60°C (Please do it in the fume hood).
- (4) Dissolve the residual with 1 mL of **Reconstitution Buffer** (Solution 3), and add 1 mL of **N-hexane**, oscillate for 30 s. Centrifuge at 4000 rpm for 5 min at room temperature.
- (5) Discard the upper liquid, take 50  $\mu$ L lower liquid for analysis.

**Note: Sample dilution factor: 2, detection limit: 3 ppb**

**3.3 Pretreatment of milk sample:**

- (1) Take 25  $\mu$ L of fresh milk sample into centrifuge tube. Add 475  $\mu$ L of **Reconstitution Buffer** (Solution 3), oscillate for 1 min and mix fully.
- (2) Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 20, detection limit: 30 ppb**

**3.4 Pretreatment of milk powder sample:**

- (1) Homogenize the sample with a homogenizer and mix fully.
- (2) Weigh  $0.5 \pm 0.05$  g of homogenized sample into 10 mL centrifuge tube, and add 5 mL of deionized water, oscillate for 1 min and mix fully.
- (3) Take 100  $\mu$ L of step (2) liquid into another centrifuge tube, and add 400  $\mu$ L of **Reconstitution Buffer** (Solution 3), oscillate for 1 min.
- (4) Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 50, detection limit: 75 ppb**

**3.5 Pretreatment of egg sample:**

- (1) Homogenize the representative sample with a homogenizer and mix fully.
- (2) Weigh  $1 \pm 0.05$  g of homogenized sample into 10 mL centrifuge tube, and add 5 mL of deionized water, oscillate for 1 min and mix fully.
- (3) Take 100  $\mu$ L of step (2) liquid into another centrifuge tube, and add 400  $\mu$ L of **Reconstitution Buffer** (Solution 3), oscillate for 1 min.
- (4) Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 30, detection limit: 45 ppb**

**3.6 Pretreatment of urine sample:**

- (1) Take 4 mL of **Reconstitution Buffer** (Solution 3) into centrifuge tube, and add 1 mL of centrifuged clear urine sample, oscillate for 30 s.
- (2) Take 50  $\mu$ L for analysis.

**Note: Sample dilution factor: 5, detection limit: 7.5 ppb**

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### 3.7 Pretreatment of serum sample:

- (1) Take 50  $\mu\text{L}$  of serum sample into centrifuge tube. Add 1450  $\mu\text{L}$  of **Reconstitution Buffer** (Solution 3), oscillate for 1 min and mix fully.
- (2) Take 50  $\mu\text{L}$  for analysis.

**Note: Sample dilution factor: 30, detection limit: 45 ppb**

### Assay procedure

Restore all reagents and samples to room temperature (25°C) before use. All the reagents should be mixed thoroughly by gently swirling before pipetting. Avoid foaming. The unused ELISA Microtiter plate should be sealed as soon as possible and stored at 2-8°C.

1. **Number:** number the sample and standard in order (multiple well), and keep a record of standard wells and sample wells. **Standard and Samples need test in duplicate.**
2. **Add sample:** add 50  $\mu\text{L}$  of **Standard or Sample** per well, then add 50  $\mu\text{L}$  of **HRP Conjugate** to each well, then add 50  $\mu\text{L}$  of **Antibody Working Solution**, cover the plate with plate sealer. oscillate for 5 s gently to mix thoroughly, incubate at 25°C for 45 min in shading light.
3. **Wash:** uncover the sealer carefully, remove the liquid in each well. Immediately add 300  $\mu\text{L}$  of **Wash Buffer** (Solution 4) to each well and wash. Repeat wash procedure for 5 times, 30 s intervals/time. Invert the plate and pat it against thick clean absorbent paper (If bubbles exist in the wells, clean tips can be used to prick them).
4. **Color Development:** add 50  $\mu\text{L}$  of **Substrate Reagent A** to each well, and then add 50  $\mu\text{L}$  of **Substrate Reagent B**. Gently oscillate for 5 s to mix thoroughly. Incubate at 25°C for 15 min in shading light (The reaction time may be shortened or prolonged according to the depth of the color).
5. **Stop reaction:** add 50  $\mu\text{L}$  of **Stop Solution** to each well, oscillate gently to mix thoroughly.
6. **OD Measurement:** determine the optical density (OD value) of each well at 450 nm (reference wavelength 630 nm) with a microplate reader. This step should be finished in 10 min after stop reaction.

## Result analysis

### 1. Absorbance(%)= $A/A_0 \times 100\%$

A: Average absorbance of standard or sample

$A_0$ : Average absorbance of 0 ppb Standard

### 2. Drawing and calculation of standard curve

Create a standard curve by plotting the absorbance percentage of each standard on the y-axis against the log concentration on the x-axis to draw a semi-logarithmic plot. Add average absorbance value of sample to standard curve to get corresponding concentration. **If samples have been diluted, the concentration calculated from the standard curve must be multiplied by the dilution factor.**

For this kit, it is more convenient to use professional analysis form for accurate and fast analysis of batch samples.

**Ofloxacin (E-FS-E129) Standard Curve**

