3th Edition, revised in Sep, 2020

（FOR RESEARCH USE ONLY. DO NOT USE IT IN CLINICAL DIAGNOSIS !）

Poly-L-lysine Solution（10×）

Catalog No：E-IR-R126

Size： 5 mL/ 10 mL

Cat.	Products	5 mL	10 mL	Storage
E-IR-R126	Poly-L-lysine Solution（10×）	5 mL	10 mL	-20°C
Manual			**One Copy**	

This manual must be read attentively and completely before using this product.

If you have any problems, please contact our Technical Service Center for help.

Phone: 240-252-7368(USA) Fax: 240-252-7376(USA)
Email: techsupport@elabscience.com
Website: www.elabscience.com

Please kindly provide us the lot number (on the outside of the box) of the kit for more efficient service.

Introduction

Poly-L-lysine Solution（PPL）is a widely used adhesive for tissue sections and glass slides. The interaction between the polycationic molecules and the anions on the tissue sections will produce a strong adhesive force. The glycoproteins on the cell surface (negative) tend to adsorb on hydrophilic surfaces, so the presence of a comparable amount of positive charge on the cell culture surface is more conducive to cell adsorption.
This product is applicable to the anti-detachment treatment of slides used in histology, immunohistochemistry, frozen section, cell smear, in situ hybridization, etc., in order to prevent tissue loss during experimental operation. It can also be used in cell culture to increase the ability of cell adhesion.

Experimental Procedure

1. Glass slide treatment
   1) The polylysine solution was diluted into 1× working solution with sterile purified water or PBS solution 9:1.
   2) Soak the slides in a diluted polylysine solution for 5 min. Note that increasing the time does not improve the package effect.
   3) Dry in the oven for 1 hour at 60 ℃ or overnight at room temperature of 18~26 ℃.
2. Treatment of culture bottle
   1) The polylysine solution was diluted into 1× working solution with sterile purified water or PBS solution 9:1.
   2) Prepare 100ml of tristeamed water for high-pressure sterilization treatment.
   3) 1×poly (lysine) working liquid was uniformly coated on the surface of the culture substrate in the amount of 50 microliters per square centimeter. It was left for 5 min at room temperature to absorb the excess liquid.
   4) Add sterilized water, rinse repeatedly for three times, and conduct aseptic operation.
   5) Sterile dry for later use.
   6) The polylysine kept aseptic after recycling can be used repeatedly.

Storage

Store at -20 ℃ for 24 months.

Cautions

1. For every 100mL diluted polylysine solution, 40~90 slides should be coated. More than 90 slides will affect the adhesion strength.
2. The slides used before must be kept clean. Wash with 70% ethanol solution containing 1% HCl if necessary.
3. The released polylysine solution should be kept at 2~8 ℃ and stable for at least 3 months.
4. Filter the used diluent and discard it if there is turbidness or long bacteria
5. The polylysine concentration can be higher, and the extracted polylysine can be reused at least 20 times.
6. For your safety and health, please wear the lab coat and disposable gloves before the experiments