



Western Blot Troubleshooting Tips

Western Blot result depends on the whole system including antigen content, sensitivity of primary antibody, sensitivity of secondary antibody, sensitivity of substrate, efficiency of color development and photographic fixing. Mistake of any procedure in the experiment may lead to unsatisfactory results.

Here Elabscience lists the common Western Blot troubleshooting.

1. No band or low band

| Possible causes | Suggestions |
|--|---|
| The loading amount of sample is low | Increase the loading amount |
| The target protein quantity is too low or not expressed in the sample | Refer to the relevant literatures to make sure that the sample contain your target protein, or prepare fresh sample again and choose a positive sample as control |
| There is a poor transfer of protein to membrane or not enough protein is bound to the membrane | Ensure that the transfer order is correct and the transfer time is sufficient |
| The antibody concentration may be too low | Use a higher concentration of antibody |
| The antibody concentration may be too high to cause the signal to disappear instantly | Use a lower concentration of antibody |
| Insufficient exposure time | Prolong the exposure time |
| Insufficient incubation or deactivation of the substrate | Increase the incubation time of substrate and ensure that the substrate is valid |
| The target protein transferred to the membrane has degraded | Keep a low transfer temperature, decrease the transfer electric current and transfer time |
| Excessive washing of the membrane | Reduce the frequency or duration of washing steps |
| The antibody is inactive or the titer of antibody is too low | Pay attention to the preservation of antibody and use antibody with higher titer |

2. High background

| Possible causes | Suggestions |
|--|---|
| The experimental equipment has been contaminated | Ensure that the equipment is clean |
| Some membrane may cause high background | NC membranes are considered to cause less background than PVDF membranes. |
| Blocking buffer is not compatible or there is cross-reactivity between the blocking buffer with antibodies | Replace the blocking buffer |
| Insufficient blocking | Prolong the blocking time |
| The antibody concentration may be too high | Use a lower concentration of antibody |
| Insufficient washing | Increase the frequency and duration of washing |
| Excessive exposure time | Shorten the exposure time |
| The membrane or buffer has been contaminated | Use the fresh buffer and keep the membrane moist during the experiment |



3. Non-specific bands

| Possible causes | Suggestions |
|--|--|
| The protein sample has digested during the treatment | Choose fresh samples for experiment |
| The loading amount of sample is too much | Reduce the loading amount |
| Insufficient blocking | Prolong the blocking time |
| The washing of membrane may be insufficient | Ensure sufficient washing of membrane |
| The antibody concentration may be too high | Use a lower concentration of antibody |
| The antibody specificity is low | Use antibody with good specificity |
| The target protein has multiple spliceosomes or modified sites | Refer to relevant literatures to check whether the target protein has other spliceosomes or modified sites |

4. Other problems

| Problems | Possible causes | Suggestions |
|---|--|--|
| Black dots on the membrane | The antibodies may have non-specifically binding with the blocking buffer | Replace the blocking buffer |
| White bands | The target protein content is too high or antibody concentration is too high | Reduce the loading amount or decrease the concentration of the antibody |
| Molecular weight is very low or high | Inappropriate gel percentage or uneven gel. /The electrophoresis temperature may be too high | Change the gel percentage: use a higher percentage for small proteins and a lower percentage for large proteins |
| Uneven bands/ Bands trail or deviate or diffuse to both sides | The equipment is not suitable. / There is bubble at the bottom/ The sample is not dissolved well. / The electrode is not balanced. / The sample amount is too much | Ensure that the electrophoresis gel is in good state and horizontal position/ Ensure the sample extraction/ Reduce the sample amount |

Summary:

The Western blot technology is rather mature, but it is not easy to get the desired result with just one trial. It is recommended to explore and optimize the experimental conditions, thus to get your ideal Western bands in the formal experiment.